## Original Contribution

# Circulating 25-Hydroxyvitamin $\mathrm{D}_{3}$ in Relation to Renal Cell Carcinoma Incidence and Survival in the EPIC Cohort 

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#### Abstract

Normal renal function is essential for vitamin D metabolism, but it is unclear whether circulating vitamin D is associated with risk of renal cell carcinoma (RCC). We assessed whether 25 -hydroxyvitamin $D_{3}\left(25(O H) D_{3}\right)$ was associated with risk of RCC and death after RCC diagnosis in the European Prospective Investigation into Cancer and Nutrition (EPIC). EPIC recruited 385,747 participants with blood samples between 1992 and 2000. The current study included 560 RCC cases, 557 individually matched controls, and 553 additional controls. Circulating $25(\mathrm{OH}) \mathrm{D}_{3}$ was assessed by mass spectrometry. Conditional and unconditional logistic regression models were used to calculate odds ratios and $95 \%$ confidence intervals. Death after RCC diagnosis was assessed using Cox proportional hazards models and flexible parametric survival models. A doubling of $25(\mathrm{OH}) \mathrm{D}_{3}$ was associated with $28 \%$ lower odds of RCC after adjustment for season of and age at blood collection, sex, and country of recruitment (odds ratio $=0.72,95 \%$ confidence interval: $0.60,0.86 ; P=0.0004$ ). This estimate was attenuated somewhat after additional adjustment for smoking status at baseline, circulating cotinine, body mass index (weight $(\mathrm{kg}) /$ height $(\mathrm{m})^{2}$ ), and alcohol intake (odds ratio $=0.82,95 \%$ confidence interval: $0.68,0.99 ; P=0.038)$. There was also some indication that both low and high $25(\mathrm{OH}) \mathrm{D}_{3}$ levels were associated with higher risk of death from any cause among RCC cases.


nested case-control study; prospective study; renal cell carcinoma; vitamin D

Abbreviations: $25(\mathrm{OH}) \mathrm{D}, 25$-hydroxyvitamin $\mathrm{D} ; 25(\mathrm{OH}) \mathrm{D}_{2}, 25$-hydroxyvitamin $\mathrm{D}_{2} ; 25(\mathrm{OH}) \mathrm{D}_{3}, 25$-hydroxyvitamin $\mathrm{D}_{3} ;$ BMI, body mass index; Cl , confidence interval; EPIC, European Prospective Investigation into Cancer and Nutrition; OR, odds ratio; RCC, renal cell carcinoma; UVB, ultraviolet B.

Vitamin D is essential for the efficient absorption of dietary calcium. Beyond its role in bone health, vitamin D has been implicated in the etiology of several cancers, most notably colorectal cancer (1,2). Vitamin D is produced in the skin after exposure to ultraviolet B (UVB) radiation from
sunlight, or it is ingested in the diet or through dietary supplements (3). After ingestion or endogenous synthesis, vita$\min \mathrm{D}$ is hydroxylated in the liver to form 25 -hydroxyvitamin D $(25(\mathrm{OH}) \mathrm{D})$, the major circulating metabolite of vitamin D, which is subsequently converted into its active form
(1,25-dihydroxyvitamin D), primarily in the kidneys. Despite the critical role of the kidneys in vitamin D metabolism, it remains unclear whether vitamin $D$ is relevant to the etiology of kidney cancer.

In 2008, the age-standardized incidence rate of kidney cancer was 3.9 cases per 100,000 people worldwide, but there is notable unexplained variation in incidence from country to country (4). A link between vitamin D and the most prevalent form of kidney cancer, renal cell carcinoma (RCC), was initially suggested on the basis of ecological evidence. For instance, ecological studies have suggested that RCC incidence is inversely associated with exposure to UVB radiation $(5,6)$. Similarly, vitamin D deficiency is highly prevalent among blacks (7), and rates of RCC are higher among blacks than whites in the United States (8).

To date, 2 prospective studies have assessed prediagnostic circulating $25(\mathrm{OH}) \mathrm{D}$ and the risk of RCC, with conflicting results $(9,10)$. Our aim was to investigate whether circulating $25(\mathrm{OH}) \mathrm{D}$ is related to the incidence of RCC and post-RCC survival using a prospective nested case-control sample from a large European cohort.

## METHODS

## Study cohort

The recruitment and baseline assessment of the European Prospective Investigation into Cancer and Nutrition (EPIC) are described in detail elsewhere (11). Between 1992 and 2000, 521,330 individuals from 10 countries were recruited, 385,747 of whom donated blood samples. Blood fractions were aliquoted into $0.5-\mathrm{mL}$ straws, which were heat sealed and stored in liquid nitrogen tanks at $-196^{\circ}$ Celsius, except in Umeå, Sweden, where samples were stored in $1.8-\mathrm{mL}$ plastic tubes in freezers at $-80^{\circ}$ Celsius and in Denmark, where samples were stored in $1-\mathrm{mL}$ plastic tubes in liquid nitrogen vapor at $-150^{\circ}$ Celsius. Participants completed self-administered questionnaires on lifestyle factors, medical history, and diet, and their heights and weights were measured using standard protocols. All participants gave written informed consent. The study was approved by the ethics committee at the International Agency for Research on Cancer (Lyon, France) and the local ethics committees of the study centers.

## Case ascertainment and follow-up

Incident cancer cases were identified via linkage to population-based cancer registries (in Denmark, Italy (except Naples), the Netherlands, Norway, Spain, Sweden, and the United Kingdom) or by active follow-up (in France, Germany, Greece, and Naples, Italy), which involved a combination of methods, including review of health insurance records and cancer and pathology registries, as well as direct contact with participants and their next-of-kin.

Mortality data were obtained from death registries at the regional or national level. Participants were followed up from study entry until cancer diagnosis (except nonmelanoma skin cancer), death, emigration, or the end of follow-up. The end of follow-up was defined as the latest date of complete followup for both cancer incidence and vital status and varied by
study center from December 2004 to June 2010. Vital status at follow-up is more than $98 \%$ complete.

## Selection of cases and controls

We identified 905 participants who were diagnosed with RCC (with International Classification of Diseases for Oncology, Second Edition, code C64.9). We excluded prevalent cases and cases with a history of another cancer (excluding nonmelanoma skin cancer, $n=85$ ); cases who did not donate a blood sample $(n=153)$; cases who had no questionnaire information available ( $n=6$ ); cases whose cancers were not histologically confirmed ( $n=27$ ); and cases from the Malmö center in Sweden, which did not participate in this study ( $n=64$ ), leaving 570 eligible RCC cases. For each case, 1 control was chosen randomly from risk sets consisting of all cohort members who were alive and free of cancer (except nonmelanoma skin cancer) at the time of diagnosis of the index case. Matching criteria were country, sex, date of blood collection ( $\pm 1$ month, which was relaxed to $\pm 5$ months for $27 \%$ of sets without available controls), and date of birth ( $\pm 1$ year, which was relaxed to $\pm 5$ years for $1 \%$ of sets without available controls). Additionally, we included 553 controls (henceforth referred to as "unmatched controls") that were individually matched to cases from another cancer site in a parallel ongoing study using identical matching criteria. These unmatched controls were included to increase the precision of the estimates.

## Biochemical analyses

Plasma samples were sent on dry ice to the Bevital AS laboratory (Bergen, Norway).Liquid chromatography coupled with tandem mass spectrometry was used to separately analyze 25-hydroxyvitamin $\mathrm{D}_{2}\left(25(\mathrm{OH}) \mathrm{D}_{2}\right)$ and 25-hydroxyvitamin $\mathrm{D}_{3}\left(25(\mathrm{OH}) \mathrm{D}_{3}\right)(12)$. The limit of detection was $3.3 \mathrm{nmol} /$ L, and within-day and between-day coefficients of variation were $4.4 \%-8.2 \%$. We found that $25(\mathrm{OH}) \mathrm{D}_{2}$ was undetectable in the majority of samples, so our analyses focus on $25(\mathrm{OH}) \mathrm{D}_{3}$. Sensitivity analyses were conducted using the sum of $25(\mathrm{OH}) \mathrm{D}_{2}$ and $25(\mathrm{OH}) \mathrm{D}_{3}$ (setting undetectable levels of $25(\mathrm{OH}) \mathrm{D}_{2}$ to 0 ), which yielded essentially identical results. Creatinine and cotinine were also assessed with liquid chromatography coupled with tandem mass spectrometry. For cotinine, the limit of detection was $1 \mathrm{nmol} / \mathrm{L}$, and the withinday and between-day coefficients of variation were $2 \%-6 \%$; for creatinine, the limit of detection was $0.25 \mu \mathrm{~mol} / \mathrm{L}$, and the within-day and between-day coefficients of variation were $2 \%-6 \%$. The laboratory is Vitamin D External Quality Assessment Scheme-certified (DEQAS, London, United Kingdom).

## Statistical analysis

We used conditional logistic regression to calculate odds ratios and $95 \%$ confidence intervals for $25(\mathrm{OH}) \mathrm{D}_{3}$ as a continuous variable, conditioning on matched case set. Concentrations were logarithmically transformed (base-2) prior to modeling, so odds ratios correspond to the expected change in odds for a doubling in concentration. We also used unconditional
logistic regression (adjusted for age at blood collection (in years), sex, and country of recruitment) to compare cases with matched controls and to the pooled group of the matched and unmatched controls. To establish whether known risk factors for RCC could explain any association, we fitted models adjusted for tobacco smoking (status at baseline of never, former, or current smoker; and quartiles of circulating cotinine determined by the distribution among current smokers), alcohol intake at recruitment (in g/day), and body mass index (BMI) (weight (kg)/height (m) ${ }^{2}$ ). As a sensitivity analysis, we fitted models that were additionally adjusted for systolic blood pressure (in mm Hg ), circulating creatinine (a marker of renal function, in $\mu \mathrm{mol} / \mathrm{L}$,), and self-reported prevalent diabetes, all of which may be on the causal pathway. We investigated potential effect modification by fitting interactions with $\log _{2} 25(\mathrm{OH}) \mathrm{D}_{3}$. Hazard ratios for all-cause mortality after RCC diagnosis were calculated using a Cox proportional hazards model with time since diagnosis as the time scale. We modeled $25(\mathrm{OH}) \mathrm{D}_{3}$ using restricted cubic splines with knots at its 10 th, 33 rd, 67 th, and 90 th percentiles. We included the same covariates as those in the unconditional logistic models, with the addition of age at diagnosis (in years). Visual inspection of smoothed, scaled Schoenfeld residuals revealed no notable departure from proportional hazards. To estimate the survival function at given concentrations of $25(\mathrm{OH}) \mathrm{D}_{3}$, we fitted a flexible parametric survival model (13), modeling the baseline cumulative hazard with restricted cubic splines (knots at the 0th, 33rd, 67th, and 100th percentiles of the distribution of failure times).

Because $25(\mathrm{OH}) \mathrm{D}_{3}$ is strongly affected by exposure to UVB radiation, all unconditional logistic models and survival models were explicitly adjusted for day of blood collection. We modeled seasonality by including 2 pairs of sine and cosine functions of day of blood collection in the models. The use of trigonometric functions adjusts for periodic variation in $25(\mathrm{OH}) \mathrm{D}_{3}$ and produces smooth predictions with no artificial discontinuities from season to season or year to year. We included 2 pairs of sine and cosine functions in the models, because the inclusion of additional terms did not improve model fit, nor did it substantially affect parameter estimates for $25(\mathrm{OH}) \mathrm{D}_{3}$.

We present 95\% confidence intervals to depict the statistical uncertainty in the estimates from the risk and survival models. We also present simulation-based estimates of statistical uncertainty, which we derived by sampling from the asymptotic distribution of the regression coefficients (the multivariate normal distribution with location and scale given by the maximum likelihood estimates and their variance-covariance matrix, respectively). We drew 1,000 samples for each model and used them to generate plausible predicted odds ratios and hazard ratios. We plotted predictions that fell within the $95 \%$ confidence interval to provide a visual impression of the $95 \%$ highest posterior density for the estimates under uniform prior distributions. $P$ values were calculated using the likelihoodratio test. The data were nearly complete for all covariates, so, where necessary, we excluded the few records with missing data. All statistical analyses were conducted using R, version 3.0.2 (R Foundation for Statistical Computing, Vienna, Austria) (14). Conditional logistic regression models were fitted using the Epi package, version 1.1.49, in R (15).

## RESULTS

## Baseline and demographic characteristics

Of the 570 matched cases and controls, 10 cases and 13 controls were missing data on $25(\mathrm{OH}) \mathrm{D}_{3}$ and were excluded from the analyses. Demographic and baseline characteristics for the remaining 560 cases, 557 matched controls, and 553 unmatched controls are presented in Table 1. The median age at diagnosis for cases was 64 years (5th and 95th percentiles, 49 and 75 years), and the median time from blood collection to diagnosis was 6.7 years ( 5 th and 95th percentiles, 0.7 and 11.9). The distributions of established risk factors for RCC showed the expected differences between cases and controls. A higher proportion of cases than controls were current smokers at baseline, and a higher proportion of cases had BMI values of 30 or higher. The unmatched controls had similar characteristics to the matched controls, albeit with a higher proportion of men and a lower proportion of participants from Sweden, Denmark, and Norway. The distribution of $25(\mathrm{OH}) \mathrm{D}_{3}$ did not vary greatly by country of recruitment (Appendix Table 1).

## Seasonal variation in plasma $25(O H) D_{3}$ concentration

There was substantial variation in plasma concentrations of $25(\mathrm{OH}) \mathrm{D}_{3}$ by date of blood collection. Figure 1 shows the observed concentrations, along with the predicted geometric mean from a linear regression model of log concentration on 2 pairs of sine and cosine functions of the day of blood collection. On average, concentrations were highest among those who had their blood drawn during or near the month of August and lowest among those who had their blood drawn during or near the months of February and March. Despite this seasonal variation, there remained substantial variability in concentration on any given day of blood collection.

## Plasma 25(OH)D ${ }_{3}$ concentration and risk of RCC

Estimated odds ratios and $95 \%$ confidence intervals for a doubling of $25(\mathrm{OH}) \mathrm{D}_{3}$ are presented in Table 2. Minimally adjusted models suggested an inverse association. Estimates from the conditional logistic model of the matched case sets were similar to those from the unconditional model (conditional odds ratio $(\mathrm{OR})=0.75,95 \%$ confidence interval (CI): $0.61,0.91, P=0.0043$; unconditional $\mathrm{OR}=0.73,95 \%$ CI: $0.59,0.89, P=0.002$ ), and the inclusion of extra unmatched controls did not substantially affect the estimated association ( $\mathrm{OR}=0.72,95 \% \mathrm{CI}: 0.60,0.86, P=0.0004$ ). Further adjustment for smoking status at baseline, circulating cotinine, alcohol intake at recruitment, and BMI attenuated the estimates somewhat, with odds ratios of $0.82(95 \% \mathrm{CI}$ : $0.68,0.99, P=0.038$ ) from the model including all controls and 0.81 ( $95 \% \mathrm{CI}: 0.65,1.00, P=0.051$ ) from the model including only matched controls. Among participants whose blood pressure was assessed at baseline ( 458 cases and 881 controls), further adjustment by continuous systolic blood pressure did not affect the estimates for $25(\mathrm{OH}) \mathrm{D}_{3}(\mathrm{OR}=$ $0.81,95 \%$ CI: $0.65,0.99$ ). Similarly, additional adjustment for prevalent diabetes did not affect the results ( $\mathrm{OR}=0.81$,

Table 1. Baseline and Demographic Characteristics of EPIC Participants, Recruited 1992-2000

| Characteristic | Cases ( $n=560$ ) |  |  |  | Matched Controls ( $n=557$ ) |  |  |  | Unmatched Controls ( $n=553$ ) |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | No. | \% | Median | Percentiles (5th, 95th) | No. | \% | Median | Percentiles (5th, 95th) | No. | \% | Median | Percentiles (5th, 95th) |
| Sex |  |  |  |  |  |  |  |  |  |  |  |  |
| Male | 311 | 56 |  |  | 309 | 55 |  |  | 374 | 68 |  |  |
| Female | 249 | 44 |  |  | 248 | 45 |  |  | 179 | 32 |  |  |
| Age at recruitment, years |  |  |  |  |  |  |  |  |  |  |  |  |
| <55 | 229 | 41 |  |  | 224 | 40 |  |  | 246 | 44 |  |  |
| 55-64.9 | 285 | 51 |  |  | 288 | 52 |  |  | 231 | 42 |  |  |
| $\geq 65$ | 46 | 8 |  |  | 45 | 8 |  |  | 76 | 14 |  |  |
| Country |  |  |  |  |  |  |  |  |  |  |  |  |
| Denmark | 114 | 20 |  |  | 114 | 20 |  |  | 0 | 0 |  |  |
| France | 13 | 2 |  |  | 13 | 2 |  |  | 7 | 1 |  |  |
| Germany | 126 | 22 |  |  | 124 | 22 |  |  | 104 | 19 |  |  |
| Greece | 17 | 3 |  |  | 17 | 3 |  |  | 22 | 4 |  |  |
| Italy | 88 | 16 |  |  | 88 | 16 |  |  | 70 | 13 |  |  |
| Netherlands | 46 | 8 |  |  | 46 | 8 |  |  | 77 | 14 |  |  |
| Norway | 4 | 1 |  |  | 4 | 1 |  |  | 2 | 0 |  |  |
| Spain | 53 | 9 |  |  | 52 | 9 |  |  | 100 | 18 |  |  |
| Sweden | 32 | 6 |  |  | 32 | 6 |  |  | 41 | 7 |  |  |
| United Kingdom | 67 | 12 |  |  | 67 | 12 |  |  | 130 | 24 |  |  |
| Smoking status at baseline |  |  |  |  |  |  |  |  |  |  |  |  |
| Never | 227 | 41 |  |  | 246 | 44 |  |  | 230 | 42 |  |  |
| Former | 162 | 29 |  |  | 179 | 32 |  |  | 199 | 36 |  |  |
| Current | 166 | 30 |  |  | 129 | 23 |  |  | 110 | 20 |  |  |
| Missing | 5 | 1 |  |  | 3 | 1 |  |  | 14 | 3 |  |  |
| Educational attainment |  |  |  |  |  |  |  |  |  |  |  |  |
| Primary school or less | 233 | 42 |  |  | 206 | 37 |  |  | 222 | 40 |  |  |
| Technical/professional school | 124 | 22 |  |  | 138 | 25 |  |  | 141 | 25 |  |  |
| Secondary school | 77 | 14 |  |  | 66 | 12 |  |  | 70 | 13 |  |  |
| Higher education | 110 | 20 |  |  | 133 | 24 |  |  | 99 | 18 |  |  |
| Missing | 16 | 3 |  |  | 14 | 3 |  |  | 21 | 4 |  |  |
| Body mass index ${ }^{\text {a }}$ |  |  |  |  |  |  |  |  |  |  |  |  |
| <25 | 182 | 32 |  |  | 223 | 40 |  |  | 220 | 40 |  |  |
| 25-29.9 | 248 | 44 |  |  | 242 | 43 |  |  | 258 | 47 |  |  |
| $\geq 30$ | 130 | 23 |  |  | 92 | 17 |  |  | 75 | 14 |  |  |
| Alcohol intake at recruitment, g/ day |  |  |  |  |  |  |  |  |  |  |  |  |
| <6 | 271 | 48 |  |  | 240 | 43 |  |  | 246 | 44 |  |  |
| 6-17.9 | 125 | 22 |  |  | 142 | 25 |  |  | 148 | 27 |  |  |
| 18-29.9 | 66 | 12 |  |  | 75 | 13 |  |  | 64 | 12 |  |  |
| $\geq 30$ | 98 | 18 |  |  | 100 | 18 |  |  | 95 | 17 |  |  |
| Age at RCC diagnosis, years |  |  | 63.8 | 48.7, 75.1 |  |  |  |  |  |  |  |  |
| Years from blood collection to diagnosis |  |  | 6.7 | $0.7,11.9$ |  |  |  |  |  |  |  |  |
| Circulating $25(\mathrm{OH}) \mathrm{D}_{3}, \mathrm{nmol} / \mathrm{L}$ |  |  | 43.2 | 17.6, 79.0 |  |  | 45.8 | 19.7, 83.2 |  |  | 48.6 | 19.5, 80.2 |
| Circulating cotinine, $\mathrm{nmol} / \mathrm{L}$ |  |  | 3 | 0, 1,703 |  |  | 2.6 | 0.0, 1,451.5 |  |  | 3.0 | 0.0, 1,500.7 |

Abbreviations: $25(\mathrm{OH}) \mathrm{D}_{3}$, 25-hydroxyvitamin $\mathrm{D}_{3}$; EPIC, European Prospective Investigation into Cancer and Nutrition; RCC, renal cell carcinoma.
${ }^{\text {a }}$ Weight (kg)/height $(\mathrm{m})^{2}$.


Figure 1. Seasonal variation of 25 -hydroxyvitamin $\mathrm{D}_{3}\left(25(\mathrm{OH}) \mathrm{D}_{3}\right)$ concentrations in plasma. Scattered points show the measured values. The solid line represents the predicted geometric mean concentration given day of blood collection, which was modeled as a linear combination of sine and cosine functions. See the text of the Methods section for further details. Estimates and data are from a renal cell carcinoma case-control sample nested within the European Prospective Investigation into Cancer and Nutrition (EPIC), which recruited participants between 1992 and 2000.
$95 \%$ CI: $0.66,0.98$ ). The estimates were also unaffected by further adjustment for circulating creatinine ( $\mathrm{OR}=0.81,95 \%$ CI: 0.66, 0.99).

Figure 2 shows the inverse association between concentration and risk by plotting odds ratio estimates from the minimally adjusted (Figure 2A) and adjusted unconditional (Figure 2B) models across the range of observed $25(\mathrm{OH}) \mathrm{D}_{3}$ concentrations. Relative to participants with a concentration of $50 \mathrm{nmol} / \mathrm{L}$, participants with less than $25 \mathrm{nmol} / \mathrm{L}$ had approximately $20 \%$ higher odds of RCC. Correspondingly, participants with concentrations greater than $100 \mathrm{nmol} / \mathrm{L} \mathrm{had}$ $20 \%$ lower odds of RCC relative to those with a concentration of $50 \mathrm{nmol} / \mathrm{L}$, but very few participants had concentrations as high as $100 \mathrm{nmol} / \mathrm{L}$.

To assess potential effect modification, we fitted models that included interaction terms between $\log _{2} 25(\mathrm{OH}) \mathrm{D}_{3}$ and various covariates. Estimates from these models are presented in Figure 3. The association with risk of RCC did not vary substantially by age at baseline, sex, country, level of education, time since blood collection, circulating concentration of creatinine, smoking status, alcohol intake at baseline, or BMI value, though there was a suggestion that the association might be slightly stronger for people with BMI values of 30 or more.

## Plasma 25(OH)D ${ }_{3}$ concentration and survival after RCC diagnosis

Of the 560 RCC cases, eight were diagnosed after the end of follow-up for vital status and were thus excluded from the survival analysis. Among the remaining 552 RCC cases, we identified 205 deaths from any cause during a median

Table 2. Odds Ratios For a Doubling in Concentration of $25(\mathrm{OH}) \mathrm{D}_{3}$ and the Risk of Renal Cell Carcinoma Among a Nested Case-Control Sample From the EPIC Cohort, Recruited 1992-2000

| Model | No. of <br> Controls | No. of <br> Cases | OR | $95 \%$ CI | $P$ Value $^{\text {a }}$ |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Minimally adjusted $^{\text {b }}$ |  |  |  |  |  |
| Conditional <br> Unconditional <br> (matched <br> controls) | 555 | 555 | 0.75 | $0.61,0.91$ | 0.0043 |
| Unconditional <br> (combined <br> controls) | 1,110 | 560 | 560 | 0.73 | $0.59,0.89$ |
| Fully adjusted |  |  | 0.002 |  |  |
| Conditional <br> Unconditional <br> (matched <br> controls) | 553 | 555 | 0.81 | $0.65,1.00$ | 0.051 |
| Unconditional <br> (combined <br> controls) | 1,092 | 555 | 0.82 | $0.68,0.99$ | 0.038 |

Abbreviations: $25(\mathrm{OH}) \mathrm{D}_{3}, 25$-hydroxyvitamin $\mathrm{D}_{3} ; \mathrm{Cl}$, confidence interval; EPIC, European Prospective Investigation into Cancer and Nutrition; OR, odds ratio.
${ }^{\text {a }} P$ values from likelihood ratio tests of $\log _{2} 25(\mathrm{OH}) \mathrm{D}_{3}$.
${ }^{\mathrm{b}}$ Conditional minimally adjusted models were conditioned on matched case set. Unconditional models were adjusted for age at baseline, country, seasonality, and sex.
${ }^{\text {c }}$ Fully adjusted models were additionally adjusted for alcohol intake at recruitment (in g/day), body mass index (weight (kg)/height (m) ${ }^{2}$ ), cotinine quartiles (based on the distribution among smokers), and smoking status at baseline (never, former, or current smoker).
follow-up of 3.24 years ( 2,397 person-years were observed in total). We found that $25(\mathrm{OH}) \mathrm{D}_{3}$ was nonlinearly associated with risk of death (likelihood ratio test of $25(\mathrm{OH}) \mathrm{D}_{3}$ terms $P=0.01$ ). Low concentrations of prediagnostic $25(\mathrm{OH}) \mathrm{D}_{3}$ were associated with increased hazards of all-cause mortality (Figure 4A). The hazard of death was 1.73 times higher ( $95 \%$ CI: 1.19, 2.51) for participants with concentrations of $25 \mathrm{nmol} / \mathrm{L}$ compared to those with concentrations of $50 \mathrm{nmol} / \mathrm{L}$. There was an indication that high concentrations might also be associated with increased hazards of death, but there were very few participants with concentrations greater than $75 \mathrm{nmol} / \mathrm{L}$. Model-based estimates of the survival function evaluated at 25, 50, and $75 \mathrm{nmol} / \mathrm{L}$ are presented in Figure 4B. The expected survival probabilities at 5 years after diagnosis were $0.56(95 \% \mathrm{CI}: 0.49,0.62)$ for participants with a concentration of $25 \mathrm{nmol} / \mathrm{L}, 0.70$ for those with a concentration of $50 \mathrm{nmol} / \mathrm{L}$, and $0.66(95 \% \mathrm{CI}: 0.58,0.73)$ for those with a concentration of $75 \mathrm{nmol} / \mathrm{L}$.

The higher hazards of death for low concentrations of $25(\mathrm{OH}) \mathrm{D}_{3}$ was apparent regardless of the time between blood collection and diagnosis; the hazard ratios for those with 25 versus $50 \mathrm{nmol} / \mathrm{L}$ were 1.75 ( $95 \% \mathrm{CI}: 1.02,2.99$ ) for those diagnosed within 5 years of blood collection and 1.67 ( $95 \% \mathrm{CI}$ : $1.02,2.71$ ) for those diagnosed 5 years or more after blood collection. In contrast, the increased hazards of death for


Figure 2. Odds ratios for renal cell carcinoma as a function of circulating concentration of 25 -hydroxyvitamin $\mathrm{D}_{3}\left(25(\mathrm{OH}) \mathrm{D}_{3}\right)$, relative to a concentration of $50 \mathrm{nmol} / \mathrm{L}$. Log-base-2 $25(\mathrm{OH}) \mathrm{D}_{3}$ was modeled as a continuous covariate. Solid and dashed lines represent the maximum likelihood estimates and $95 \%$ confidence intervals, respectively. The translucent lines are 1,000 draws from the multivariate normal distribution defined by the maximum likelihood estimates and their variance-covariance matrix; they thus give an indication of the posterior density for the odds ratio under a uniform prior on the regression coefficients. The "rug plot" under each panel shows the observed distribution of 25 -hydroxyvitamin $D_{3}$. Estimates and data are from a nested case-control sample within the European Prospective Investigation into Cancer and Nutrition (EPIC), which recruited participants between 1992 and 2000. A) Estimates adjusted for age at baseline, sex, country, and seasonality (sine and cosine functions of day of blood collection). B) Estimates after additional adjustment for smoking status at baseline (never/former/ current smoker), circulating cotinine (quartiles defined among the controls), alcohol intake at recruitment (in g/day), and body mass index (weight $(\mathrm{kg}) /$ height $\left.(\mathrm{m})^{2}\right)$.
higher concentrations was apparent only among those diagnosed within 5 years of blood collection, with hazard ratios for 75 versus $50 \mathrm{nmol} / \mathrm{L}$ of 2.22 ( $95 \% \mathrm{CI}: 1.49,3.28$ ) for those diagnosed within 5 years of blood collection and 0.62 ( $95 \% \mathrm{CI}: 0.33,1.15$ ) for those diagnosed more than 5 years after blood collection (likelihood ratio test of interaction terms $P=0.001$ ). Estimated hazard ratios did not vary by sex, age, country, educational level, alcohol intake, smoking status, or circulating creatinine (data not shown).

## DISCUSSION

We found suggestive evidence that circulating concentrations of $25(\mathrm{OH}) \mathrm{D}_{3}$ are inversely associated with the risk of RCC, such that participants with concentrations of less than $25 \mathrm{nmol} / \mathrm{L}$ had approximately $20 \%$ greater risk than those with concentrations of $50 \mathrm{nmol} / \mathrm{L}$. We also found that lower concentrations of $25(\mathrm{OH}) \mathrm{D}_{3}$ were nonlinearly associated with the risk of all-cause mortality after RCC diagnosis. Among the majority of participants, an inverse association was apparent, whereas higher concentrations of $25(\mathrm{OH}) \mathrm{D}_{3}$ also appeared to be associated with higher risk of death.

Previous reports of prospectively measured circulating $25(\mathrm{OH}) \mathrm{D}$ and the risk of kidney cancer have produced conflicting results. In accordance with our results, the Copenhagen City Heart Study, a cohort of 9,791 people, including 55 incident kidney cancer cases, reported that a $50 \%$ reduction in $25(\mathrm{OH}) \mathrm{D}$ was associated with higher risk (hazard ratio $=$
$1.34,95 \%$ CI: 1.04, 1.73) (9). In contrast, the Vitamin D Pooling Project found no association in an analysis of 775 case-control pairs nested within 8 prospective cohorts (10). These discrepant results are not readily explicable. One difference between the Vitamin D Pooling Project and the present study is the method of adjustment for season. The Vitamin D Pooling Project used conditional logistic regression models adjusted for season of blood collection (summer vs. winter), with sensitivity analyses adjusted for seasonality by using the residuals from a local polynomial regression (16). In contrast, we directly modeled seasonality using smooth trigonometric functions. Another difference between the studies is that both the Vitamin D Pooling Project and the Copenhagen City Heart Study used a chemiluminescence immunoassay measuring both $25(\mathrm{OH}) \mathrm{D}_{2}$ and $25(\mathrm{OH}) \mathrm{D}_{3}$, whereas in the present study, we used liquid chromatography coupled with tandem mass spectrometry to quantify $25(\mathrm{OH}) \mathrm{D}_{3}$ specifically. That said, these methodological differences would seem unlikely to fully account for the discrepant results, which remain unexplained.

Although few studies have directly assessed vitamin D status, some investigators have taken a different approach, creating a predicted vitamin D score on the basis of established determinants of vitamin $D$ concentrations $(17,18)$. Joh et al. (19) investigated predicted $25(\mathrm{OH}) \mathrm{D}$ concentrations (calculated on the basis of race, UVB flux, physical activity, BMI value, vitamin D intake, alcohol consumption, and postmenopausal hormone use) and risk of RCC among participants of the Nurses' Health Study and the Health Professionals


Figure 3. Stratified odds ratios (ORs) and $95 \%$ confidence intervals (CIs) for renal cell carcinoma for a doubling in concentration of 25 -hydroxyvitamin $D_{3}$. Estimates are adjusted for age at baseline, sex, country, seasonality (sine and cosine functions of day of blood collection), smoking status at baseline (never/former/current smoker), circulating cotinine (quartiles defined among the controls), alcohol intake at recruitment (in g/day), and body mass index (BMI) (weight (kg)/height $\left.(\mathrm{m})^{2}\right)$. P values are from likelihood ratio tests of interaction terms. Estimates are from a nested case-control sample within the European Prospective Investigation into Cancer and Nutrition (EPIC), which recruited participants between 1992 and 2000. Bars, 95\% Cls.

Follow-up Study. They found a strong inverse association between predicted $25(\mathrm{OH})$ D and risk, such that a $10-\mathrm{ng} / \mathrm{mL}$ (approximately $25-\mathrm{nmol} / \mathrm{L}$ ) increment in predicted score was associated with a $44 \%$ lower hazard of RCC. Although the magnitude of the estimated association is greater, this result is broadly consistent with our observation that incrementing $25(\mathrm{OH}) \mathrm{D}_{3}$ from 25 to $50 \mathrm{nmol} / \mathrm{L}$ is associated with approximately $20 \%$ lower risk. In contrast, studies of dietary sources of vitamin D alone have largely yielded null results (20-22), possibly because dietary sources do not contribute greatly to circulating vitamin D concentrations (17).

There are several plausible mechanisms that might underpin an association between vitamin D and RCC (23). For
instance, it is possible that vitamin D modifies the effects of risk factors such as obesity, hypertension, or diabetes. Although we observed an indication that the association between $25(\mathrm{OH}) \mathrm{D}_{3}$ and RCC risk might be slightly stronger among those with BMI values of 30 of higher, statistical adjustment for systolic blood pressure or prevalent diabetes did not affect the estimates, suggesting a potential role of vitamin D beyond that of established risk factors. This is consistent with studies of human RCC cell lines and murine RCC, which have shown that vitamin D species can inhibit tumor cell proliferation, angiogenesis, and metastasis $(24,25)$. Conversely, given the critical role of the kidneys in vitamin D metabolism, it is possible that the observed association is driven


Figure 4. Post-renal cell carcinoma ( RCC ) survival. Estimates from a nested case-control sample within the European Prospective Investigation into Cancer and Nutrition (EPIC), which recruited participants between 1992 and 2000. A) Hazard ratios from a Cox model for all-cause mortality after RCC diagnosis as a function of circulating concentration of 25 -hydroxyvitamin $D_{3}\left(25(\mathrm{OH}) \mathrm{D}_{3}\right)$, relative to a concentration of $50 \mathrm{nmol} / \mathrm{L}$. We modeled $25(\mathrm{OH}) \mathrm{D}_{3}$ using restricted cubic splines with knots at the 10 th, 33 rd, 67 th, and 90 th percentiles of its distribution. The model was adjusted for age at baseline, sex, country, and seasonality (sine and cosine functions of day of blood collection), smoking status at baseline (never/former/ current smoker), circulating cotinine (quartiles defined among the controls), alcohol intake at recruitment (in g/day), and body mass index (weight $(\mathrm{kg}) /$ height $\left.(\mathrm{m})^{2}\right)$. Solid and dashed lines represent the maximum likelihood estimates and $95 \%$ confidence intervals, respectively. The translucent lines are 1,000 draws from the multivariate normal distribution defined by the maximum likelihood estimates and their variance-covariance matrix; they thus give an indication of the posterior density for the hazard ratio under a uniform prior on the regression coefficients. The "rug plot" shows the observed distribution of $25(\mathrm{OH}) \mathrm{D}_{3}$. B) Survival function after RCC diagnosis evaluated at given concentrations of $25(\mathrm{OH}) \mathrm{D}_{3}$, derived from a flexible parametric survival model. Restricted cubic splines with knots at the 0 th, $33 \mathrm{rd}, 67 \mathrm{th}$, and 100th percentiles of the distribution of uncensored survival times were used to model the baseline hazard. We modeled $25(\mathrm{OH}) \mathrm{D}_{3}$ using restricted cubic splines with knots at the 10th, 33 rd, 67 th, and 90 th percentiles of its distribution.
by perturbed vitamin D metabolism as a consequence of early kidney dysfunction. Although the association remained consistent throughout follow-up and was not affected by adjustment for circulating creatinine, we cannot completely rule out the possibility that early renal dysfunction was the cause, rather than the result, of the observed distribution of circulating vita$\min$ D.

Many researchers have investigated circulating vitamin D and all-cause mortality in general populations. Consistent with our observation, many studies have reported higher risk of death for people with low vitamin D concentrations (2633). This suggests that the association observed in our study may not be specific to RCC survival, but rather a reflection of a general phenomenon. Our observation that high levels of $25(\mathrm{OH}) \mathrm{D}_{3}$ might be associated with higher risk of death is consistent with results from the Uppsala Longitudinal Study of Adult Men, which also suggest a U-shaped association (34). Despite this accord, further studies are required to investigate the intriguing possibility that both low and high concentrations are associated with all-cause mortality.

The principal limitation of our study is that $25(\mathrm{OH}) \mathrm{D}_{3}$ was measured using a single blood sample drawn in adulthood. Although individual vitamin D measurements are reasonably reproducible, intraindividual variation may still be important
(35). Further, it is possible that a single measurement in adulthood does not capture exposure to vitamin D in an etiologically relevant period.

Our study has several strengths. Importantly, our sample included participants from 10 European countries from different geographical latitudes and with a wide range of $25(\mathrm{OH}) \mathrm{D}_{3}$ concentrations. Biospecimen handling was standardized, and quantification of circulating $25(\mathrm{OH}) \mathrm{D}_{3}$ took place in a single laboratory, thus minimizing systematic interlaboratory variation. The prospective design of our study, in which $25(\mathrm{OH}) \mathrm{D}_{3}$ concentrations were assessed using blood collected prior to diagnosis, minimizes the chance that any differences between cases and controls are caused by existing tumors. Further, the availability of detailed information on potential confounders-particularly the inclusion of circulating cotinine as a biomarker of current smoking intensity and creatinine as a marker of renal function-affords additional confidence that the observed associations were not caused by residual confounding.

In conclusion, we found that low concentrations of $25(\mathrm{OH}) \mathrm{D}_{3}$ were associated with higher risk of RCC as well as lower all-cause mortality among RCC cases. High concentrations of $25(\mathrm{OH}) \mathrm{D}_{3}$ might also be associated with increased risk of all-cause mortality among RCC cases.

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## REFERENCES

1. Jenab M, Bueno-de Mesquita HB, Ferrari P, et al. Association between pre-diagnostic circulating vitamin D concentration and risk of colorectal cancer in European populations: a nested case-control study. BMJ. 2010;340:b5500.
2. Touvier M, Chan DS, Lau R, et al. Meta-analyses of vitamin D intake, 25-hydroxyvitamin D status, vitamin D receptor
polymorphisms, and colorectal cancer risk. Cancer Epidemiol Biomarkers Prev. 2011;20(5):1003-1016.
3. Holick MF. McCollum Award Lecture, 1994: vitamin D-new horizons for the 21st century. Am J Clin Nutr. 1994;60(4): 619-630.
4. Ferlay J, Shin HR, Bray F, et al. Estimates of worldwide burden of cancer in 2008: GLOBOCAN 2008. Int J Cancer. 2010; 127(12):2893-2917.
5. Grant WB, Garland CF. The association of solar ultraviolet B (UVB) with reducing risk of cancer: multifactorial ecologic analysis of geographic variation in age-adjusted cancer mortality rates. Anticancer Res. 2006;26(4A):2687-2699.
6. Mohr SB, Gorham ED, Garland CF, et al. Are low ultraviolet B and high animal protein intake associated with risk of renal cancer? Int J Cancer. 2006;119(11):2705-2709.
7. Ginde AA, Liu MC, Camargo CA Jr, et al. Demographic differences and trends of vitamin D insufficiency in the US population, 1988-2004. Arch Intern Med. 2009;169(6):626-632.
8. Lipworth L, McLaughlin JK, Tarone RE, et al. Renal cancer paradox: higher incidence but not higher mortality among African-Americans. Eur J Cancer Prev. 2011;20(4):331-333.
9. Afzal S, Bojesen SE, Nordestgaard BG. Low plasma 25-hydroxyvitamin D and risk of tobacco-related cancer. Clin Chem. 2013;59(5):771-780.
10. Gallicchio L, Moore LE, Stevens VL, et al. Circulating 25-hydroxyvitamin D and risk of kidney cancer: Cohort Consortium Vitamin D Pooling Project of Rarer Cancers. Am J Epidemiol. 2010;172(1):47-57.
11. Riboli E, Hunt KJ, Slimani N, et al. European Prospective Investigation into Cancer and Nutrition (EPIC): study populations and data collection. Public Health Nutr. 2002; 5(6B):1113-1124.
12. Midttun $\emptyset$, Ueland PM. Determination of vitamins A, D and E in a small volume of human plasma by a high-throughput method based on liquid chromatography/tandem mass spectrometry. Rapid Commun Mass Spectrom. 2011;25(14): 1942-1948.
13. Royston P, Parmar MK. Flexible parametric proportionalhazards and proportional-odds models for censored survival data, with application to prognostic modelling and estimation of treatment effects. Stat Med. 2002;21(15):2175-2197.
14. R Core Team. R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing: Vienna, Austria; 2013.
15. Carstensen B, Plummer M, Laara E, et al. Epi: A package for statistical analysis in epidemiology. R package version 1.1.49. 2013.
16. Gallicchio L, Helzlsouer KJ, Chow WH, et al. Circulating 25-hydroxyvitamin D and the risk of rarer cancers: design and methods of the Cohort Consortium Vitamin D Pooling Project of Rarer Cancers. Am J Epidemiol. 2010;172(1):10-20.
17. Bertrand KA, Giovannucci E, Liu Y, et al. Determinants of plasma 25-hydroxyvitamin D and development of prediction models in three US cohorts. Br J Nutr. 2012;108(10): 1889-1896.
18. Giovannucci E, Liu Y, Rimm EB, et al. Prospective study of predictors of vitamin D status and cancer incidence and mortality in men. J Natl Cancer Inst. 2006;98(7):451-459.
19. Joh HK, Giovannucci EL, Bertrand KA, et al. Predicted plasma 25 -hydroxyvitamin D and risk of renal cell cancer. J Natl Cancer Inst. 2013;105(10):726-732.
20. Bosetti C, Scotti L, Maso LD, et al. Micronutrients and the risk of renal cell cancer: a case-control study from Italy. Int J Cancer. 2007;120(4):892-896.
21. Prineas RJ, Folsom AR, Zhang ZM, et al. Nutrition and other risk factors for renal cell carcinoma in postmenopausal women. Epidemiology. 1997;8(1):31-36.
22. Wilson RT, Wang J, Chinchilli V, et al. Fish, vitamin D, and flavonoids in relation to renal cell cancer among smokers. Am J Epidemiol. 2009;170(6):717-729.
23. Giovannucci E. The epidemiology of vitamin D and cancer incidence and mortality: a review (United States). Cancer Causes Control. 2005;16(2):83-95.
24. Fujioka T, Hasegawa M, Ishikura K, et al. Inhibition of tumor growth and angiogenesis by vitamin $\mathrm{D}_{3}$ agents in murine renal cell carcinoma. J Urol. 1998;160(1):247-251.
25. Nagakura K, Abe E, Suda T, et al. Inhibitory effect of 1 alpha,25-dihydroxyvitamin $\mathrm{D}_{3}$ on the growth of the renal carcinoma cell line. Kidney Int. 1986;29(4):834-840.
26. Schöttker B, Haug U, Schomburg L, et al. Strong associations of 25-hydroxyvitamin D concentrations with all-cause, cardiovascular, cancer, and respiratory disease mortality in a large cohort study. Am J Clin Nutr. 2013;97(4):782-793.
27. Virtanen JK, Nurmi T, Voutilainen S, et al. Association of serum 25 -hydroxyvitamin D with the risk of death in a general older population in Finland. Eur J Nutr. 2011;50(5): 305-312.
28. Hutchinson MS, Grimnes G, Joakimsen RM, et al. Low serum 25 -hydroxyvitamin D levels are associated with increased all-cause mortality risk in a general population: the Troms $\varnothing$ Study. Eur J Endocrinol. 2010;162(5):935-942.
29. Szulc P, Claustrat B, Delmas PD. Serum concentrations of $17 \beta-\mathrm{E} 2$ and 25 -hydroxycholecalciferol (25OHD) in relation to all-cause mortality in older men-the MINOS Study. Clin Endocrinol (Oxf). 2009;71(4):594-602.
30. Semba RD, Houston DK, Ferrucci L, et al. Low serum 25 -hydroxyvitamin D concentrations are associated with greater all-cause mortality in older community-dwelling women. Nutr Res. 2009;29(8):525-530.
31. Pilz S, Dobnig H, Nijpels G, et al. Vitamin D and mortality in older men and women. Clin Endocrinol (Oxf). 2009;71(5): 666-672.
32. Melamed ML, Michos ED, Post W, et al. 25-hydroxyvitamin D levels and the risk of mortality in the general population. Arch Intern Med. 2008;168(15):1629-1637.
33. Jia X, Aucott LS, McNeill G. Nutritional status and subsequent all-cause mortality in men and women aged 75 years or over living in the community. Br J Nutr. 2007;98(3):593-599.
34. Michaëlsson K, Baron JA, Snellman G, et al. Plasma vitamin D and mortality in older men: a community-based prospective cohort study. Am J Clin Nutr. 2010;92(4):841-848.
35. Major JM, Graubard BI, Dodd KW, et al. Variability and reproducibility of circulating vitamin D in a nationwide U.S. population. J Clin Endocrinol Metab. 2013;98(1):97-104.

Appendix Table 1. Distribution of $25(\mathrm{OH}) \mathrm{D}_{3}$ by Country of
Recruitment Among Renal Cell Carcinoma Cases and Controls
Nested Within EPIC, Recruitment 1992-2000

| Country | No. | Percentile of $\mathbf{2 5 ( O H ) \mathrm { D } _ { 3 } , \text { nmol/L }}$ |  |  |
| :--- | ---: | ---: | ---: | :---: |
|  |  | $\mathbf{5 \%}$ | $\mathbf{5 0 \%}$ | $\mathbf{9 5 \%}$ |
| Denmark | 228 | 18.19 | 50.68 | 89.11 |
| France | 33 | 19.30 | 45.63 | 86.47 |
| Germany | 354 | 18.01 | 41.72 | 78.00 |
| Greece | 56 | 18.72 | 42.16 | 65.90 |
| Italy | 246 | 14.55 | 40.08 | 77.00 |
| Netherlands | 169 | 22.54 | 45.79 | 78.91 |
| Norway | 10 | 25.06 | 52.42 | 71.83 |
| Spain | 205 | 22.37 | 43.97 | 77.84 |
| Sweden | 105 | 33.92 | 57.07 | 85.34 |
| United Kingdom | 264 | 24.25 | 51.26 | 82.73 |
| $\quad$ Overall | 1,670 | 18.90 | 45.69 | 81.31 |

Abbreviations: $25(\mathrm{OH}) \mathrm{D}_{3}$, 25 -hydroxyvitamin $\mathrm{D}_{3}$; EPIC, European Prospective Investigation into Cancer and Nutrition.

